# THE INFLUENCE OF CERTAIN TREATMENTS WITH PHYSICAL MUTAGENE AGENTS ON THE ACTIVITY OF SOME ENZYMES IN ECHINACEA PURPUREA AND HYPERICUM PERFORATUM

## ELENA CIORNEA<sup>1\*</sup>, V. ARTENIE<sup>1</sup>, DANIELA ICHIM<sup>2</sup>, G. GHEORGHITA<sup>2</sup>, D.COJOCARU<sup>1</sup>, DANIELA VARARU<sup>1</sup>

Keywords: catalase, peroxidase, gamma radiation, Echinacea purpurea, Hypericum perforatum

Abstract: We determined catalase and peroxidase activity in *Echinacea purpurea* and *Hypericum perforatum* plantlets resulted after the germination of seeds irradiated with radiation doses of 1, 3, 5, 8, 10, 12, 15 and 20 KRad, the radiation flow being of 0.5 KRad per minute. After interpreting the results obtained we found the radiation influence on enzyme activity, in the respect of its stimulation at low radiation doses.

#### **INTRODUCTION**

The catalase (E.C. 1.11.1.6) and peroxidase (E.C. 1.11.1.7) are di-component enzymes that belong to the haemoprotein class, having the hemine as prosthetic group, that is the IX ferri-porphyrine.(Artenie and Tanase,1981).

These oxido-reducing enzymes have a significant biological role, being involved in the protection against stressful factors, such as radiation, pesticides a.s.o.(Corneanu,1989). In the specialised literature there are lots of data on the influence of radiation over vegetal organisms, in the respect of causing oxidative stress, as suite of formation of reactive oxygen species.(Cojocaru,1997)

#### Purpose of the research

This works aims to obtain certain data regarding the dynamics of certain enzymes' activity in the *Echinacea purpurea* and *Hypericum perforatum* plantlets, after their treatment with gamma radiation at different radiation doses.

This is furthermore justified by the well-known fact that these herbs are extremely appreciated for their therapeutic qualities, among which the antiviral effect shows up by itself, envisaging the acquire of immune-stimulating, anti-inflammatory, scarring and anti-HIV drugs. (Ciulei et al, 1993).

### MATERIAL AND METHODS

The research was carried out on 14-days plantlets obtained from irradiated seeds that were put to germinate in controlled conditions.

The biological samples come from the Secuieni Agro-Zootechnical Research Facility, Neamt County, and the seeds were irradiated at the Nuclear Unity of ICCF Bucharest with increased radiation doses, at the flow of 0.5 KRad/min.

The catalase activity has been determined by iodine titration.(Artenie and Tanase,1981) and the peroxidase activity by the colorimetric method with o-dianisidine(Möler and Ottolenghi ,1966).

The extraction of catalase from the biological material was made by  $Na_2HPO_4$  solution 0.1M, and that of the peroxidase with phosphate buffer 0.4M.

We carried out 4 to 6 determinations for each sample and witness separately, and the results we obtained were statistically processed by using the STUDENT test.

### **RESULTS AND DISCUSSIONS**

The experiments performed have outlined the influence of radiation on the enzyme activity in the analysed samples.

r	<u>`</u>	<u> </u>			
Sample	Ν	Average	Standard	Probability	
			error		
М	4	3.5646	±0.1948	-	
P1	4	3.6120	±0.0490	p>0.5	
P3	4	4.0156	±0.1063	0.5>p>0.25	
P5	4	5.8100	±0.1722	0.01>p>0.002	
P8	4	5.4750	±0.0333	0.002>p>0.001	
P10	4	2.9746	±0.1917	0.5>p>0.25	
P15	4	6.4600	±0.1294	0.01>p>0.002	
P20	4	6.2046	±0.0649	0.002>p>0.001	

Table I: Catalase activity(CU/g/min) in Echinacea purpurea:

Legend: M = non-irradiated witness

P1 = sample with 1 KRad radiation dose

P3 = sample with 3 KRad radiation dose

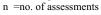
P5 = sample with 5 KRad radiation dose

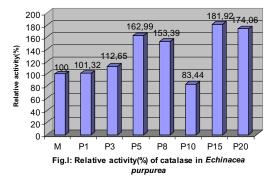
P8 = sample with 8 KRad radiation dose

P10 = sample with 10 KRad radiation dose

P15 = sample with 15 KRad radiation dose

P20 = sample with 20 KRad radiation dose



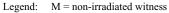


## Legend: idem Table I

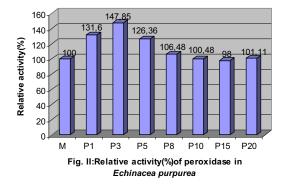
As one can see, at low radiation doses (1, 3 KRad respectively), the activity is similar to that of the witness, while in the P5 and P8 samples one can see a strong stimulation of the enzyme activity.

	~ _	- 0 /		
Sample	n	Average	Standard	Probability
			error	
M	4	2.0058	$\pm 0.0744$	-
P1	4	2.6398	$\pm 0.0856$	P<0.001
P3	4	2.9656	±0.0919	P<0.001
P5	4	2.5346	$\pm 0.0848$	0.5>P>0.25
P8	4	2.1358	±0.1356	0.01 <p<0.002< td=""></p<0.002<>
P10	4	2.0156	$\pm 0.0185$	0.05>P>0.02
P15	4	1.9658	$\pm 0.0068$	0.25>P>0.1
P20	4	2.0282	±0.1256	0.01>P>0.002

Table II:Peroxidase activity(PU/g/min) in Echinacea purpurea:



P1 = sample with 1 KRad radiation dose P3 = sample with 3 KRad radiation dose P5 = sample with 5 KRad radiation dose P8 = sample with 8 KRad radiation dose P10 = sample with 10 KRad radiation dose P15 = sample with 15 KRad radiation dose P20 = sample with 20 KRad radiation dose n =no.of assessments.



Legend: idem Table II.

The peroxidase activity for the species of *Echinacea purpurea* recorded a significant increase at low radiation doses (1 and 3 KRad), opposite to the catalase, while at high doses the activity becomes similar to that of the witness.

 use dett (i) (ee, gillin) in Hypertetint perjor ditunt.					
Sample	n	Average	Standard	Probability	
			error		
М	4	0.7556	±0.0929	-	
P1	4	1.3173	±0.0185	0.05>p>0.02	
P3	4	1.0270	±0.0069	0.25>p>0.1	
P5	4	0.9663	±0.0251	0.5>p>0.25	
P8	4	0.9423	±0.0241	0.5>p>0.25	
P12	4	1.2446	±0.0184	0.05>p>0.02	
P15	4	0.3223	±0.0121	0.1>p>0.05	

Table III:Catalase activity(CU/g/min) in *Hypericum perforatum*:

Legend:

M = non-irradiated witness

P1 = sample with 1 KRad radiation dose

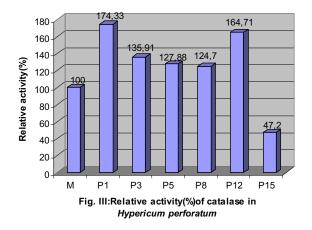
P3 = sample with 3 KRad radiation dose

P5 = sample with 5 KRad radiation dose

P8 =sample with 8 KRad radiation dose

P12 = sample with 12 KRad radiation dose P15 = sample with 15 KRad radiation dose

n =no.of assessments



#### Legend: idem Table III

From the above data we find the catalase activity was strongly stimulated by small radiation doses, whilst at 15 KRad the enzyme's activity was strongly inhibited, thus recording a much lesser value compared to the witness.

Table IV:Peroxidase activity(PU/g/min) in *Hypericum perforatum*:

Sample	n	Average	Standard	Probability
			error	
М	6	4.2085	±0.0944	-
P1	6	6.5912	±0.1022	p<0.001
P3	6	7.6755	±0.2651	p<0.001
P5	6	3.0757	$\pm 0.0848$	0.01 <p<0.002< td=""></p<0.002<>
P8	6	4.4526	±0.1383	p<0.5
P12	6	4.1466	±0.0937	p>0.5
P15	6	6.1818	±0.2226	0.05 <p<0.02< td=""></p<0.02<>

Legend:

M = non-irradiated witness P1 = sample with 1 KRad radiation dose

P3 = sample with 3 KRad radiation dose

P5 = sample with 5 KRad radiation dose

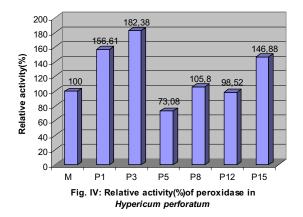
P8 =sample with 8 KRad radiation dose

P12 =sample with 0 KRad radiation dose

 $P_{12}$  - sample with 12 KRad radiation dose  $P_{15}$  = sample with 15 KRad radiation dose

n =no.of assessments

Subsequent to the research undertaken on *Hypericum perforatum*, we found that peroxidase activity was strongly stimulated at doses of 1, 3 KRad respectively, inhibited at 5 KRad, whilst high doses of 15 KRad caused a new increase in enzyme's activity.



Legend: idem Table IV.

#### CONCLUSIONS

Low radiation doses caused an important increase in peroxidase activity in *Echinacea purpurea*, while the catalase activity in the same species recorded a significant increase at doses of 5, 8, 15 and 20 KRad respectively.

In the species *Hypericum perforatum*, we found a stimulation of peroxidase activity at 1, 3 KRad, while catalase activity is increased at low radiation doses and strongly inhibited at 15 KRad.

The results obtained are similar to those in the specialized literature, low radiation doses causing a stimulation of enzyme activity.

### REFERENCES

Artenie Vlad, Tănase Elvira,1981. Practicum de biochimie generală, Centrul de multiplicare al Univ.Al.I.Cuza, Iași, p. 135;

**Ciulei I., Grigorescu E., Stănescu Ursula, 1993**. Plante medicinale-Fitochimie și Fitoterapie, vol.II, Ed. Medicală, București, p.11; 690;

Cojocaru D., 1997. Enzimologie, Ed. Gama, Iași, p. 88;

**Corneanu G, 1989.** Radiosensibilitatea plantelor sub impactul factorilor stresanți- principii de radiologie vegetală, Ed. Ceres, București, , p. 45;

**Gheorghiță G.I., 1996-1997.** Efectele dozelor slabe de iradiere la plante (radio-stimulare), Stațiunea "Stejarul", Pângărați, p. 139;

**Möler K.M.and Ottolenghi P., 1966.** The oxidation of o-dianisidine by  $H_2O_2$  and peroxidase at neutral pH, vol. 35, Compts. Rend. Tray. Lab., Carlsberg, p. 369;

**Păun E., Mihalea A., Anela Dumitrescu , Maria Verzea, Oltea Coșocariu, 1986.** Tratat de plante medicinale și aromatice, Tome I, Ed. Academiei Republicii Socialiste Romane., București, p. 318.

<sup>1</sup>)Al. I. Cuza University, Iasi.

<sup>2</sup>)G. Bacovia University, Bacău,

\* ciornea@email.ro